

# A REVIEW ON SEMISOLID TOPICAL PREPARATION

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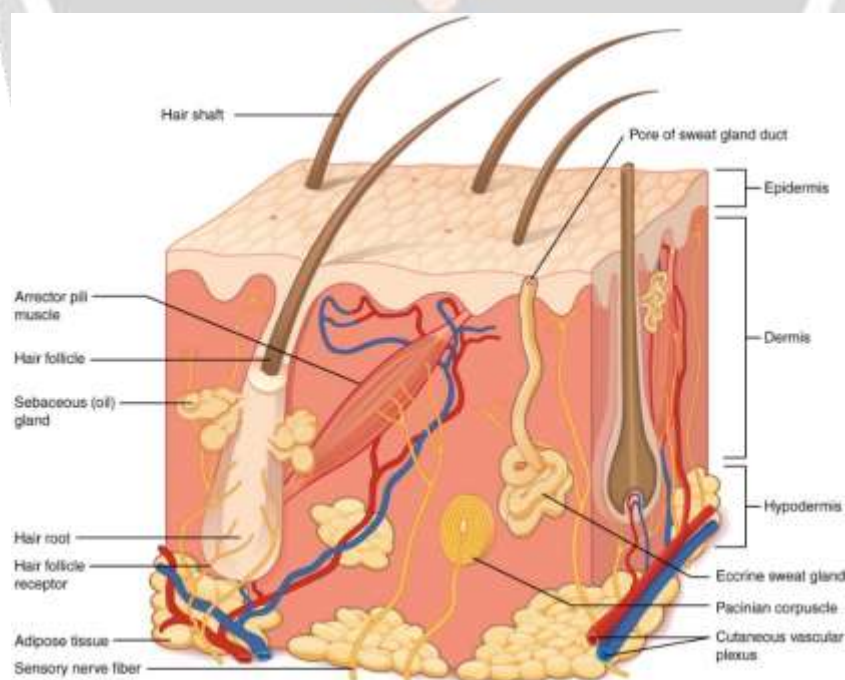
## Abstract

*Already in ancient times, semisolid preparations for cutaneous application, popularly known as ointments, played an important role in human society. An advanced scientific investigation of “ointments” as dosage forms was set off in the late fifties of the previous century. It was only from then on that the intensive physico-chemical characterization of ointments as well as the inclusion of dermatological aspects led to a comprehensive understanding of the various interactions between the vehicle, the active ingredient, and the skin. In the meantime, many researchers have been involved in optimizing semisolid formulations with respect to continuously changing therapeutic and patient needs. Aspects that have been dealt with are the optimization of dermato-biopharmaceutical properties and many different issues related to patient’s compliance, such as skin tolerance, applicability, and cosmetic appeal. Moreover, processing technology has been improved and analytical techniques developed and refined in order to enable improved characterization of the formulation itself as well as its interaction with the skin.*

**Keywords:** - semi-solid dosage form, dermatology products

## 1. INTRODUCTION

Skin is the most important and has a complex structure. It is the largest and easily accessible organ of living animals, however here the discussion is limited to human beings.<sup>1,2</sup> It is estimated that an average adult human has a skin surface area of 1.8 to 2.0 m<sup>2</sup>. Morphologically skin forms the most complex structure comprising of multiple layers with thickness ranging from 0.5 to 4 mm.<sup>3,4</sup> Hence, understanding the skin morphology becomes essential before discussing skin diseases, route of administration and permeation/ penetration of chemicals (active or inactive) through skin.<sup>5,6</sup>



**Figure 1.** Diagrammatic representation of layers of Skin.

Skin has three layers majorly viz. epidermis, dermis and hypodermis.<sup>7</sup> Stratum corneum forms the outer most layer of epidermis which forms the major barrier (rate limiting layer) for most of the actives as well as foreign particulate matter including microorganism to enter through skin.<sup>8,9</sup> Dermis and hypodermis have numerous blood vessels and consists of hair follicles, sweat glands and other structures like sebaceous glands.<sup>10</sup>

Topical dosage forms are solutions, suspensions, creams, gels, ointments or pastes containing one or more active ingredients dissolved or uniformly dispersed in a suitable base.<sup>11,12</sup> Depending on the disease condition and desired therapeutic effect the dosage form design is selected.<sup>13</sup> The excipient selection depends largely on dosage form being developed, which forms the most critical part of development in Topical dosage form as it will decide the characteristics of end-product which is directly impacting active ingredient efficacy and product elegance.<sup>14,15</sup>

With increase in understanding on skin morphology and route of penetration, interest on skin formulation (Topical Semisolid Dosage form) has tremendously increased in pharmaceutical Research over the last few decades.<sup>16,17</sup> Conventional emulsions, creams ointments and lotions are still acceptable and cost-effective option available for skin disorders.<sup>18</sup>

Indian Pharmaceutical product manufacturers contribute around 30% to 35% of generic market globally due to potential research scope and low conversion cost. Various pharmaceutical sectors are effortlessly developing, approving and manufacturing Generic Drug Products for global supply.<sup>19,20,21</sup> Generic Drug Products are bioequivalent cost-effective replacement for Branded / Patented formulations which could be used as alternative with equivalent therapeutic effectiveness. While discussing development of generic Topical Semisolid dosage form; selection of excipients, its quantity, grade, selection of right vendor as well as manufacturing process used to produce Topical Products are considered as critical attributes for success rate of formulation.<sup>22,23,24</sup> Topical products are considered to be complex products where the therapeutic effect depends on drug transport from product to skin surface and from skin surface to site of action, hence the micro-arrangement of excipients and actives within the formulation.<sup>25,26,27</sup>

Semisolids are the product for the use on skin and mucous membranes. It may used for the treatment of pathological conditions and may also protect the body from harmful conditions. These semisolid preparations are used to protect over the time and produces the therapeutic effect through the occlusions.<sup>28,29,30</sup> These may contain the plastic rheological behaviour. These behaviours may allow the semisolid to retain the shape and size of the film and act by the outside force. For localized drug delivery these semi solid preparations are use.<sup>31,32</sup> In recent era, these semisolid preparations are used for the systemic delivery of various types of drugs. These give the significance in pharmaceutical dosage forms. It may be applied on the skin, cornea, rectal tissue, nasal mucosa, vagina and external ear lining.<sup>33,34,35</sup>

These preparations may contain two phases. One is external phase and another one is internal phases. These active ingredients are present in both the phases.<sup>36,37</sup> Three-dimensional structure may characterize the semisolids preparation.<sup>38</sup>

### **Ideal Properties of Semisolid Dosage Forms**

#### **1. Physical Properties**

- Smooth texture
- Elegant in appearance
- Non dehydrating
- Non gritty, greasy and non-staining

#### **2. Physiological Properties**

- Non irritating
- Do not alter membrane / skin functioning
- Miscible with skin secretion
- low sensitization index

#### **3. Application Properties**

- Easily applicable with efficient drug release
- High aqueous washability

#### **4. Storage Properties**

- Storage of semisolids at 25°C.
- It should not allow to freeze and stored in a well-closed container.<sup>39,40,41,42</sup>

## 2. FACTORS AFFECTING SKIN PENETRATION

- gastro intestinal absorption and skin penetration both are same by the rate of diffusion, pH, physicochemical property, vehicle used and concentration.
- In physicochemical factor the skin penetration is hydration state of stratum corneum. It may affect the rate of penetration of skin.<sup>43,44,45</sup>
- to the absorption site, solubility of a drug determines the concentration. The rate of transport influences the partition coefficient.
- In absorption rate and the molecular weight an inverse relationship is appeared.
- large molecule penetrates slow then small molecules. There is a co relation between the size and penetration rate due to molecular size.<sup>46,47,48</sup>

## 3. TOPICAL DRUG DELIVERY

An Intestinal lipid pathway and proteinaceous cellular component is present in between the layer of skin and stratum corneum.<sup>49,50</sup> By the help of tortuous and intercellular path drug molecules are penetrated into the skin. By the help of solvents and enhancers are used for the transport of topical drugs.<sup>51,52</sup> It may also transport by the drug by the hair follicles, transcellular and sweat ducts. The molecules which are presents are only penetrate through the skin.<sup>53,54</sup> Ointment may retain the significance of trans epidermal of water and drug transport by the hydrated skin. dermatitis, psoriasis, and warts are the barrier of skins.<sup>56,57</sup> These are the geared of drugs towards skin diseases. Driving forces are generated between the formulation and site of action it may help the drug to penetrated into the skin.<sup>58,59</sup> A thermodynamically activity may generate large driving forces. It helps to transport the formulation through the skin at a small amount of fraction.<sup>60,61</sup> In super-saturated conditions the thermodynamically activity has a greater unity and may enhanced the drugs through skin. It is in metastable state and it may convert back into their original sizes with changing in the flux of drug.<sup>62,63,64</sup>

## 4. MATERIALS USED FOR MEDICINAL PREPARATION

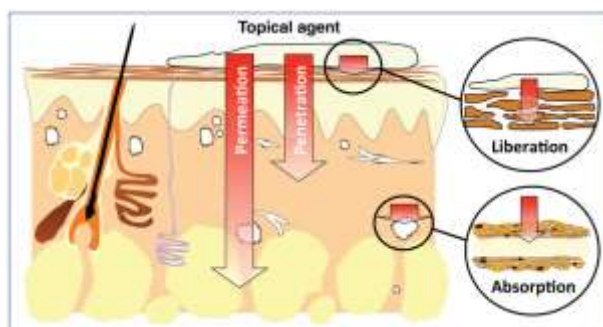
Physiochemical properties like solubility, molecular weight, pH and stability are shown for the composition of medicinal preparation.<sup>65,66</sup> The ingredients which are required for the manufacturing of medicinal preparation are been given by according to pharmaceutical law, patent law. Packaging and stability are also considered. Solvents are also used in this.<sup>67,68</sup> Vehicles like preservatives, antioxidants, dyes, and buffers are also used for this preparation. No means trivial may have the composition of suitable, well-balanced, and stable preparation.<sup>69,70,71</sup>

## 5. QUALITY REQUIRED FOR MEDICINAL PREPARATION

Medicinal preparation is required a quality criteria. Quality is the main factor for this preparation.<sup>72,73</sup> This are also help for the development of medicinal products. Every cosmetic and medical device required validated and manufactured according to high quality standards. All ingredients should be characterized by high purity and quality.<sup>74,75,76</sup> These manufactures required certification and quality assurance processes. Each final batch should undergo testing like chemical, physical and microbiological stability. Stress test are performed for the determination of in-use and storage stability. AMG is also applied for every formulation.<sup>77,78,79</sup>

## 6. PHARMACOKINETIC OF TROPICAL AGENTS

Tropical agent's pharmacokinetics were determined by physiochemical characteristics of active ingredients. It may also depend on vehicles characteristics and the skin condition.<sup>80,81</sup> Different pharmaceutical concepts have different vehicles system and these may affect the pharmacokinetics of medicinal preparation. Skin conditions are also paly a most important role. The preparation selection is based on pharmacokinetic criteria.<sup>82,84,85</sup>



**Figure 2.** Pharmacokinetics of topical agents

Pharmacokinetic principle may have three overlapping functioning process. The epicutaneous of tropical agents may dissolved and suspended the ingredient released.<sup>83,84</sup> Only this preparation may become the skin bioavailability. Vehicle composition may liberate the rate of physicochemical properties as well as properties of skin.<sup>85,86</sup> The skin may be inter and intra which may be changes the healing process. The therapeutics effects may also varies. The stratum corneum has play an important role as boundary layer and a drug uptake.<sup>87,88</sup> This stratum corneum has reservoir function. Semi solid preparations may transfer the stratum corneum within a few minutes before residue removed from the skin.<sup>89,90</sup> Active ingredients contain numerous microcompartments at the skin layers. Some factors may impact this process. diffusion distance, diffusion surface and concentration differences are the most important in this process.<sup>91,92</sup> Hagen-Poiseuille's law of hydrodynamic pressure gradient show the thermodynamic activity and convection processes.<sup>93,94</sup> The active ingredient may pass through the skin layer and may diffused into deeper tissue layers. These may be eliminated by absorption through the microvascular system or by enzymatic degradation by sessile cell.<sup>95,96,97</sup> For diffusion process various routes is available. Intercellular route is the most important passage marked as hydrophilic and a lipophilic route. This utilizes the hydrophilic of ceramides and water molecule interacting for diffusion.<sup>98,99,100</sup> For matrix penetration, ceramides have flexible lipophilic side chains. dermatological indication and pharmacodynamic properties of an active ingredient may used for the optimization of concentration-time profile.<sup>101,102</sup> Indication based approach is optimized the therapeutic efficacy. The diseased skin contains chemical and physical factors.<sup>103</sup>

## 7. COMBINATION PREPARATION AND PLANNING OF TREATMENT

For clinical treatment, some combination of active pharmaceutical ingredient. Physicochemical properties in active ingredient may be optimized.<sup>104</sup> Combination preparation were pharmaceutical represented. pharmaceutical ingredients of two ingredient may combined and have clinical efficacy to improve the patient treatment.<sup>105,106</sup> antifungal agent and corticosteroid combination may have inflammatory cutaneous mycoses. combination therapy has extemporaneous formulations in medicinal products.<sup>107,108</sup> Pharmaceutical incompatibilities and interactions are not trivial. On skin lesion the polyvalent preparation were applied without clearing the surface.<sup>109,110</sup> Other therapeutic has antibiotic/antiseptic agents, corticosteroids and local anaesthetic agents has not complete the requirement of the treatment.<sup>111,112</sup> through specific and nonspecific adverse effects pose and inefficient risk is undue. Mono preparation of sequential are used in the combination of therapeutic. 30minute interval is shown it reduces the risk of interaction in between the ointment and the skin.<sup>113,114</sup>

For optimization of therapeutic intervention are able to order by medical, economic and aspect of practical.<sup>115</sup> These are used in severe and chronic disease. Disease acuteness, induction and maintenance of therapy may prove the successfulness. Pathogenesis of disease may have therapeutic components.<sup>116,117</sup> These processes may contain physical, cosmetic, pharmacological and physiotherapeutic. For tropical applications in medicinal products suitable vehicles are used and the dosage form may produce the important role.<sup>118,119</sup> Only one concentration is used into the active ingredient. These medicaments are having some chances in curative treatment.<sup>120,121</sup>

## 8. ADVANCED MEDICAL DEVICES USED IN THE TREATMENT OF SKIN DISEASES

There are three possibilities by which semi solid preparation may applied on the skin. These are tropical drugs, cosmetics and medical devices.<sup>122,123</sup> Drugs may be defined as a substances or mixer of substances which are used for the treatment or administration to heal and prevent diseases.<sup>124,125</sup> It may also used for the correction of pharmacological, immunological and metabolic means.<sup>126</sup>

Medical devices are the substances which are used for the prevention, monitoring, treatment and alleviation of host from the body.<sup>127</sup> It is not done by pharmacological or immunological means or by metabolic means.

Medical devices are different form any other process by the legal basis. A compressive licence is required for the drug supply to the markets.<sup>128,129</sup>

### **Atopic dermatitis**

It is a skin inflammatory chronic disease. It may affect the children. It may characterise by xerosis, erythematous and pruritus lesions with trans epidermal. Some microorganism is also used in this skin infections.<sup>130,131</sup> one semi solid preparation is used for the management of atopic dermatitis. For the reduction of signs and symptoms of atopic dermatitis nonsteroidal multicomponent are used. corticosteroid and calcineurin are used in multicomponent treatment for distinguished from other topical preparations. Atopic dermatitis is the treatment which may treat mild to moderate diseases in child and adult.<sup>132,133</sup>



### **X-ray- or radiation-induced dermatitis**

In cancer therapy, radiotherapy is most important treatment. Dose depended skin reaction may receive radiation induced dermatitis.<sup>134</sup> These may be ringing from mild erythema to moist desquamation. double blind and vehicle controlled the efficacy and tolerability of medical devices.<sup>135,136</sup>



### **Seborrheic dermatitis**

This skin disease manifests the scalp and the face and is associated with scaling.<sup>137</sup> Antifungal agents and calcineurin inhibitors are the medical device and may reduce the seborrheic dermatitis.<sup>138</sup>



### **Actinic keratosis**

It may be defined as a scaly, erythematous lesions produce by exposure of sun for many years. It was only found by sun terraces on the skin. It was the neoplasias in the fair skinned population and may produces nonmelanoma skin cancer.<sup>139,140</sup> In situ it was consider as a squamous cell carcinoma. It may transform into squamous cell carcinoma. These may require early treatment. It may be characterized by hyperkeratotic, scaling, lesions by sun exposed skin sites. UV radiation may produce the nonmelanoma skin cancer.<sup>141,142</sup> This preparation is already established for the prevention of epithelial cutaneous lesions. Curettage, cryotherapy, excision and laser therapy is used for the treatment and prevention of squamous cell carcinomas.<sup>143,144</sup> These lesions have greater importance. Liposomal sunscreen is used as a medical device in that.<sup>145,146</sup>



### Hypertrophic scars and keloids

For the treatment of scars corticosteroids and silicones are used. Silicone gels are easy to handle.<sup>147</sup> The mechanism of action of silicone has hydration of stratum corneum by which the trans epidermal water loss is reduced.<sup>148</sup> Cytokine signal transformation from keratinocytes to dermal fibroblasts. It may inhibit the proliferation of fibroblast and should reduce the collagen synthesis.<sup>149,150</sup> Self-driving silicon gels are used for the prevention and treatment of hypertrophic scars and keloids in medical devices. Silicon gels has lower height.<sup>151</sup>

### REFERENCES

1. Wohlrab J. Grundlagen der topischen Therapie. *Hautarzt* 2014; 65: 169–74.
2. Staubach P, Lunter DJ. Basistherapie in der Dermatologie – Geeignete Grundlagen, Möglichkeiten und Grenzen. *Hautarzt* 2014; 65: 63–72.
3. Wohlrab J, Klauck D, Savtcheva E. Regulatorische Besonderheiten für Topika. *Hautarzt* 2014; 65: 175–9.
4. Mills PC, Magnusson BM, Cross SE. The effects of vehicle and region of application on in vitro penetration of testosterone through canine skin. *Vet J* 2006; 171: 276–80.
5. Seidel K. Rezepturen konservieren. *DAZ* 2015; 43: 59–65.
6. Silva P. The right skin preparation technique: a literature review. *J Perioper Pract* 2014; 24: 283–5.
7. Symposium der Fachgruppe Arzneimittelkontrolle/Pharmazeutische Analytik der Deutschen Pharmazeutischen Gesellschaft (DPHG). Reinheitsforderungen an Arzneimittel – Notwendigkeiten und Grenzen. *Pharmazie in unserer Zeit* 1997; 26: 88–97.
8. Schaefer H, Jamouille JC. Skin pharmacokinetics. *Int J Dermatol* 1988; 27: 351–9.
9. Sennhenn B, Giese K, Plamann K et al. In vivo evaluation of the penetration of topically applied drugs into human skin by spectroscopic methods. *Skin Pharmacol Physiol* 1993; 6: 152–60.

10. Wohlrab J. Stellenwert der Therapieplanung. *Hautarzt* 2014; 65: 218–20.
11. Barbosa CD, Balp MM, Kulich K et al. A literature review to explore the link between treatment satisfaction and adherence, compliance, and persistence. *Patient Preference Adherence* 2012; 6: 39–48.
12. Swarbrick J, Boylan J. C., *Encyclopedia of Pharmaceutical Technology*. Vol. 14, 1996. Marcel Dekker Inc. 31-59.
13. Lachman L, Lieberman H. A, Kanig J. L., *Theory and Practice of Industrial Pharmacy*. 4<sup>th</sup> Indian Edition. 1991. Verghese Publishing House. 534-63.
14. Jani G. K., *Dispensing Pharmacy*. 3rd Edition. 2003-04. B.S. Shah Publication. 201 -03, 22.
15. Banker G. S., Rhodes C.T., *Modern Pharmaceutics*. Vol. 7. 1979. Marcel Dekker Inc. 272-76.
16. Chater S.J., *Cooper and Gunn Dispensing Pharmacy For Pharmaceutical Students*. 12th Edition. 2001. CBS Publication. 192-231
17. Aulton M. E., *Pharmaceutics the Science of Dosage Form Design: 1st Edition*. 1995. ELBS Churchill Livingstone. 386.
18. Barry B. W., *Dermatological Formulations*. Vol. 18. 1983. Marcel Dekker Inc. 296-340
19. Gupta P., Garg S., Recent Advances in Semisolid Dosage Form for Dermatological Application. *Pharmaceutical Technology*. March 2002. 144 -62
20. Basu S, Chakraborty S, Bandyopadhyay AK, Development and Evaluation of a Mucoadhesive Nasal Gel of Midazolam Prepared with *Linum usitatissimum* L. Seed Mucilage, *Scientia Pharmaceutica*, 2009; 77: 899–910.
21. Alsarra A et al., Mucoadhesive Polymeric Hydrogels for Nasal Delivery of Acyclovir, *Drug Development and Industrial Pharmacy*, 2009; 35:352–62.
22. Varshosaz J, Sadrai H, Heidari A, Nasal Delivery of Insulin Using Bioadhesive Chitosan Gels, *Drug Delivery*, 2006; 13:31–6.
23. Pisal SS, Reddy P, Paradkar AR, Mahadik KR, Kadam SS, Nasal Melatonin gels using Pluronic PF-127 for chronobiological treatment of sleep disorder, *Indian Journal of Biotechnology*, July 2004; 3:369-77.
24. Majithiya RJ, Ghosh PK, Umrethia ML, Murthy RSR., Thermoreversible-mucoadhesive Gel for Nasal Delivery of Sumatriptan. *AAPS PharmSciTech*. 2006; 7(3): Article 67.
25. Nandgude T, Thube R, Jaiswal N, Deshmukh P, Chatap V, Hire N, Formulation and Evaluation of pH Induced In-situ Nasal Gel of Salbutamol Sulphate, *International Journal of Pharmaceutical Sciences and Nanotechnology*, July –September 2008; 1(2):177-83.
26. Mahajan HS, Gattani S, In situ gels of Metoclopramide Hydrochloride for intranasal delivery: In vitro evaluation and in vivo pharmacokinetic study in rabbits, *Drug Delivery*, 2010; 17(1): 19–27.
27. Rathnam G, Narayanan N, Ilavarasan R, Preparation And Evaluation Of Carbopol Based Nasal Gels For Systemic Delivery Of Progesterone, *International Journal Of Periodontics and Restorative Dentistry*, March 2005; 2(1).
28. Hussain, et al. Nasal dosage forms of propranolol, United States Patent-4394390.
29. Kuotsu K, Bandyopadhyay A, Development of Oxytocin Nasal Gel using Natural Mucoadhesive Agent obtained from the Fruits of *Dellinia indica* L., *Science Asia*, 2007; 33: 57-60.
30. Alexandrou TJ et al., Reduction of preoperative conjunctival bacterial flora with the use of mupirocin nasal ointment, *Trans Am Ophthalmol Soc.*, 2006; 104:196-201.
31. Giandalia G, Caro V, Cordone L, Giannola LI, Trehalose hydroxyethylcellulose microspheres containing vancomycin for topical drug delivery, *European Journal of Pharmaceutics and Biopharmaceutics*, 2001; 52: 83-9.
32. Tas C, Ozkan Y, Savaser A, Baykara T, In vitro release studies of chlorpheniramine maleate from gels prepared by different cellulose derivatives, *Il Farmaco*, 2003; 58 : 605-11.
33. Kang L. et al, SMGA gels for the skin permeation of haloperidol, *Journal of Controlled Release*, 2005; 106: 88–98.
34. Wu Z, Controlled release of lidocaine hydrochloride from the surfactant-doped hybrid xerogels, *Journal of Controlled Release*, 2005; 104: 497–505.
35. Valenta C, Nowack E, Deoxycholate-hydrogels: novel drug carrier systems for topical use, *International Journal of Pharmaceutics*, 1999; 185:103–11.
36. Patel M. et al, Preparation and characterization of oxybenzone loaded gelatin microspheres for enhancement of sunscreen efficiency, *Drug Delivery*, 2006; 13: 323-30.
37. Dinarvand R, Rahmani E, Farbod E, Gelatin Microspheres for the Controlled Release of All-trans-Retinoic Acid Topical Formulation and Drug Delivery Evaluation, *Iranian Journal of Pharmaceutical Research*, 2003; 47-50.
38. Gogna D et al, Microsphere based improved sunscreen formulation of ethylhexyl methoxycinnamate, *Current Drug Delivery*, 2007 Apr; 4(2):153-9.

39. Yoshida H. et al, In vitro release of Tacrolimus from Tacrolimus ointment and its speculated mechanism, *International Journal of Pharmaceutics*, 2004;270:55–64.
40. Doijad RC et al, Sustained ophthalmic delivery of Gatifloxacin from in situ gelling system, *Indian Journal of Pharmaceutical Science*, 2006; 814-5.
41. Pandit JK et al, Long-acting ophthalmic formulation of indomethacin: evaluation of alginate gel system, *Indian journal of pharmaceutical science*, 2007; 37-8.
42. Sechoy O et al, A new long-acting ophthalmic formulation of Carteolol containing alginate acid, *International Journal of Pharmaceutics*, 2000;207 :109–16.
43. Kaur IP, Singh M, Kanwar M, Formulation and evaluation of ophthalmic preparations of acetazolamide, *International Journal of Pharmaceutics*, 2000;199: 119–27.
44. Srividya B. et al, sustained ophthalmic delivery of ofloxacin from a pH triggered in situ gelling system, *Journal of Controlled Release*, 2001; 73: 205–11.
45. Qi H. et al, Development of a poloxamer analogs/carbopol-based in situ gelling and mucoadhesive ophthalmic delivery system for puerarin, *International Journal of Pharmaceutics*, 2007;337:178–187.
46. Wagh VD et al, Formulation and Evaluation of Ophthalmic Insert Drug Delivery System of Forskolin, *Asian journal of Pharmaceutics*, 221-4.
47. Gupta H. et al, pH-Induced in Situ Gel for Periodontal Anesthesia, *Indian Journal of Pharmaceutical Science*, 2009.
48. Bhowmik BB et al, Formulation Development and Characterization of Metronidazole Microencapsulated Bioadhesive Vaginal Gel, *International Journal of Pharmacy and Pharmaceutical Sciences*, 2009; 1(1): 240-57.
49. Ramadan AA, Formulation and Evaluation of Bioadhesive Gels Containing Miconazole Nitrate, *Journal of Applied Sciences Research*, 2008; 4(9): 1052-65.
50. Kumar J. et al, Formulation of Thermoresponsive and Buccal Adhesive In Situ Gel for Treatment of Oral Thrush containing Itraconazole, *Journal of Pharmaceutical Sciences & Research*, 2010;2(2): 116-22.
51. Shin SC, Bum JP, Choi JS, Enhanced bioavailability by buccal administration of triamcinolone acetonide from the bioadhesive gels in rabbits, *International Journal of Pharmaceutics*, 2009; 209: 37–43.
52. Yong SC et al, Enhanced rectal bioavailability of ibuprofen in rats by poloxamer 188 and menthol, *International Journal of Pharmaceutics*, 2004; 269: 169–76.
53. Kubo W. et al, Oral sustained delivery of ambroxol from in situ-gelling pectin formulations, *International Journal of Pharmaceutics*, 2004; 271:233–40.
54. Harish NM et al, Formulation and Evaluation of in situ Gels Containing Clotrimazole for Oral Candidiasis, *Indian Journal of Pharmaceutical Sciences*, 2009; 421-3
55. Rajnikanth PS, Mishra B, Floating in situ gelling system for stomach site-specific delivery of clarithromycin to eradicate *H. pylori*, *Journal of Controlled Release*, 2008; 125:33–41.
56. Ricci EJ et al, Sustained release of lidocaine from Poloxamer 407 gels, *International Journal of Pharmaceutics*, 2005; 288: 235–44.
57. Guo DD et al, Synergistic anti-tumor activity of paclitaxel-incorporated conjugated linoleic acid-coupled poloxamer thermosensitive hydrogel in vitro and in vivo, *Biomaterials*, 2009; 30: 4777–85.
58. Schorn G. *MPG Medizinproduktegesetz, 4. Auflage*, Wissenschaftliche Verlagsgesellschaft, Stuttgart, 2009.
59. Platzeck T, Krätke R, Schulz C. Kosmetische Mittel – Sicherheitsaspekte. *Bundesgesundheitsblatt* 2010; 53: 610–4.
60. Rogiers V, Pauwels M. Cosmetic products and their current European regulatory framework. *Curr Probl Dermatol* 2008; 36: 1–28.
61. Schlemminger M. The proof of the pudding – die Zulassung. In: Dagmar Fischer, Jörg Breitenbach (Hrsg.): *Die Pharmaindustrie. 2. Auflage*, Heidelberg: Spektrum Akademischer Verlag, 2010.
62. Korting HC. Evidenzbasierte Dermatika und Kosmetika. In: Plewig G, Kaudewitz P, Sander CA (Hrsg): *Fortschritte der praktischen Dermatologie und Venerologie. Band 19*, Berlin: Springer, 2004: 548–54.
63. Pavicic T, Steckmeier S, Kerscher M, Korting HC. Evidenzbasierte Kosmetika: Konzepte und Anwendung bei den Zielstellungen Cellulite, Striae distensae und Acne vulgaris. *Geburtsh Frauenheilk* 2007;68: 986–9.
64. Korting HC, Borelli C, Schöllmann C. Acne vulgaris. Rolle der Kosmetik. *Hautarzt* 2010; 61: 126–31.
65. Getting medical devices faster to market. *Med Device Technol* 2007; 18: 52.



66. Patrizi A, Raone B, Neri I. Atopiclair. *Expert Opin Pharmacother* 2009; 10: 1223–30.
67. Hanifin JM, Rajka G. Diagnostic features of atopic dermatitis. *Acta Derm Venereol (Stockh)* 1980; 92(Suppl.): 44–7.
68. Cork MJ, Danby SG, Vasilopoulos Y, Hadgraft J, Lane E, Moustafa M, Guy RH, MacGowan AL, Tazi-Ahni R, Ward SJ. Epidermal barrier dysfunction in atopic dermatitis. *J Invest Dermatol* 2009; 129: 1892–908.
69. Boguniewicz M, Leung DYM. Recent insights into atopic dermatitis and implications for management of infectious complications. *J Allergy Clin Immunol* 2010; 125: 4–13.
70. Abramovits W, Hebert AA, Boguniewicz M, Kempers SE, Tschen E, Jarratt MT, Lucky AW, Cornelison RL, Swinyer LJ, Jones TM. Patient-reported outcomes from a multicenter, randomized, vehicle-controlled clinical study of MAS063DP (Atopiclair) in the management of mild-to-moderate atopic dermatitis in adults. *J Dermatolog Treat* 2008; 19: 327–32.
71. Belloni G, Pinelli S, Veraldi S. A randomised, double-blind, vehicle-controlled study to evaluate the efficacy and safety of MAS063D (Atopiclair) in the treatment of mild to moderate atopic dermatitis. *Eur J Dermatol* 2005 ;15: 31–6.
72. Patrizi A, Capitanio B, Neri I, Giacomini F, Sinagra JL, Raone B, Berardesca E. A double-blind, randomized, vehicle-controlled clinical study to evaluate the efficacy and safety of MAS063DP (ATOPICLAIR) in the management of atopic dermatitis in paediatric patients. *Pediatr Allergy Immunol* 2008; 19: 619–25.
73. Boguniewicz M, Zeichner JA, Eichenfield LF, Hebert AA, Jarratt M, Lucky AW, Paller AS. MAS063DP is effective monotherapy for mild to moderate atopic dermatitis in infants and children: a multicentre, randomized, vehicle-controlled study. *J Pediatr* 2008; 152: 854–9.
74. King KB, Nail LM, Kreamer K, Strohl RA, Johnson JE. Patients' descriptions of the experience of receiving radiation therapy. *Oncol Nurs Forum* 1985; 12: 55–61.
75. Leonardi MC, Gariboldi S, Ivaldi GB, Ferrari A, Serafini F, Didier F, Mariani L, Castiglioni S, Orecchia R. A doubleblind, randomised, vehicle-controlled clinical study to evaluate the efficacy of MAS065D in limiting the effects of radiation on the skin: interim analysis. *Eur J Dermatol* 2008; 18: 317–21.
76. Primavera G, Carrera M, Berardesca E, Pinnaró P, Messina M, Arcangeli G. A double-blind, vehicle-controlled clinical study to evaluate the efficacy of MAS065D (Xclair), a hyaluronic acidbased formulation, in the management of radiation-induced dermatitis. *Cutan Ocul Toxicol* 2006; 25: 165–71.
77. Veraldi S, Menter A, Innocenti M. Treatment of mild to moderate seborrhoeic dermatitis with MAS064D (Sebclair), a novel topical medical device: results of a pilot, randomized, doubleblind, controlled trial. *J Eur Acad Dermatol Venereol* 2008; 22: 290–6.
78. Rőwert-Huber J, Patel MJ, Forschner T, Ulrich C, Eberle J, Kerl H, Sterry W, Stockfleth E. Actinic keratosis is an early in situ squamous cell carcinoma: a proposal for reclassification. *Br J Dermatol* 2007; 156(Suppl 3): 8–12.
79. Moy RL. Clinical presentation of actinic keratoses and squamous cell carcinoma. *J Am Acad Dermatol* 2000;42: 8–10.
80. Wlaschek M, Tancheva-Poór I, Naderi L, Ma W, Schneider LA, Razi-Wolf Z, Schüller J, Scharffetter-Kochanek K. Solar UV irradiation and dermal photoaging. *J Photochem Photobiol B* 2001; 63: 41–51.
81. Terhorst D, Stockfleth E. Therapie epithelialer Tumoren. *Akt Dermatol* 2008; 34: 482–6.
82. Ulrich M, Drecoll U, Stockfleth E. Emerging drugs for actinic keratosis. *Expert Opin Emerg Drugs* 2010; 15: 545–55.
83. Ulrich C, Degen A, Patel MJ, Stockfleth E. Sunscreens in organ transplant patients. *Nephrol Dial Transplant* 2008; 23: 1805–8.
84. Roberts RJ, Burgess IF. New head-lice treatments: hope or hype? *Lancet* 2005;365: 8–9.
85. Richter J, Müller-Stöver I, Walter S, Mehlhorn H, Häussinger, D. Kopfläuse – Umgang mit einer wieder auflebenden Parasitose. *Dtsch Arztebl* 2005; 102: A-2395/B-2016/C-1909.
86. Mehlhorn H (Hrsg.). *Encyclopedic references of parasitology*. 1. Auflage, Springer Verlag, Berlin, 2002: 339–41.
87. Richling I, Böckeler W. Lethal Effects of Treatment with a Special Dimeticone Formula on Head Lice and House Crickets (Orthoptera, Ensifera: *Acheta domestica* and Anoplura, Phthiraptera: *Pediculus*

- humanus*) Arzneimittel-Forschung 2008; 58: 248–54.
88. Oliveira FA, Speare R, Heukelbach J. High in vitro efficacy of Nyda L, a pediculicide containing dimeticone. *J Eur Acad Dermatol Venereol* 2007; 21: 1325–9.
  89. Heukelbach J, Asenov A, Liesenfeld O, Mirmohammadsadegh A, Oliveira FA. A new two-phase dimeticone pediculicide shows high efficacy in a comparative bioassay. *BMC Dermatol* 2009; 9: 12.
  90. Sonnberg S, Oliveira FA, de Melo ILA, de Melo Soares MM, Becher H, Heukelbach J. Ovizide Wirksamkeit von over-the-counter Kopflausprodukten. Poster10-33-17 (Postersession 38- 34-4), präsentiert bei der 104. Jahrestagung der Deutschen Gesellschaft für Kinder- und Jugendmedizin in München 2008.
  91. Heukelbach J, Pilger D, Oliveira FA, Khakban A, Ariza L, Feldmeier H. A highly efficacious pediculicide based on dimeticone: randomized observer blinded comparative trial. *BMC Infect Dis* 2008; 8: 115.
  92. Mustoe TA, Cooter RD, Gold MH, Hobbs FD, Ramelet AA, Shakespeare PG, Stella M, Téot L, Wood FM, Ziegler UE; International Advisory Panel on Scar Management. International clinical recommendations on scar management. *Plast Reconstr Surg* 2002; 110: 560–71.
  93. Mustoe TA. Evolution of silicone therapy and mechanism of action in scar management. *Aesthetic Plast Surg* 2008; 32: 82–92.
  94. Chernoff WG, Cramer H, Su-Huang S. The efficacy of topical silicone gel elastomers in the treatment of hypertrophic scars, keloid scars, and post-laser exfoliation erythema. *Aesthetic Plast Surg* 2007; 31: 495–500.
  95. Signorini M, Clementoni MT. Clinical evaluation of a new self-drying silicone gel in the treatment of scars: a preliminary report. *Aesthetic Plast Surg* 2007; 31: 183–7.
  96. Spellberg, B.; Bartlett, J.G.; Gilbert, D.N. The future of antibiotics and resistance. *N. Engl. J. Med.* 2013, 368, 299–302.
  97. Aryee, A.; Price, N. Antimicrobial stewardship—Can we afford to do without it? *Br. J. Clin. Pharmacol.* 2015, 79, 173–181.
  98. Cosgrove, S.E.; Carmeli, Y. The impact of antimicrobial resistance on health and economic outcomes. *Clin. Infect. Dis.* 2003, 36, 1433–1437.
  99. Neu, H.C. The crisis in antibiotic resistance. *Science* 1992, 257, 1064–1073.
  100. Levy, S.B.; Marshall, B. Antibacterial resistance worldwide: Causes, challenges and responses. *Nat. Med.* 2004, 10, 122–129.
  101. Schüts, E.C.; Hulscher, M.E.; Mouton, J.W.; Verduin, C.M.; Stuart, J.W.; Overdiek, H.W.; van der Linden, P.D.; Natsch, S.; Hertogh, C.M.; Wolfs, T.F.; et al. Current evidence on hospital antimicrobial stewardship objectives: A systematic review and meta-analysis. *Lancet Infect. Dis.* 2016, 16, 847–856.
  102. Sulakvelidze, A.; Alavidze, Z.; Morris, J.G., Jr. Bacteriophage therapy. *Antimicrob. Agents Chemother.* 2001, 45, 649–659.
  103. Cisek, A.A.; Dabrowska, I.; Gregorczyk, K.P.; Wyzewski, Z. Phage therapy in bacterial infections treatment: One hundred years after the discovery of bacteriophages. *Curr. Microbiol.* 2017, 74, 277–283.
  104. Bruynoghe, R.; Maisin, J. Essais de therapeutique au moyen du bacteriophage du staphylocoque. *J. Compt. Rend Soc. Biol.* 1921, 85, 1020–1121.
  105. Chanishvili, N. Phage therapy-history from twort and d'herelle through soviet experience to current approaches. *Adv. Virus Res.* 2012, 83, 3–40.
  106. Debarbieux, L.; Pirnay, J.P.; Verbeken, G.; De Vos, D.; Merabishvili, M.; Huys, I.; Patey, O.; Schoonjans, D.; Vanechoutte, M.; Zizi, M.; et al. A bacteriophage journey at the european medicines agency. *Microbiol. Lett.* 2016, 363.
  107. Vandenheuvel, D.; Lavigne, R.; Brussow, H. Bacteriophage therapy: Advances in formulation strategies and human clinical trials. *Annu. Rev. Virol.* 2015, 2, 599–618.
  108. Loc-Carrillo, C.; Abedon, S.T. Pros and cons of phage therapy. *Bacteriophage* 2011, 1, 111–114.
  109. Kutter, E.; De Vos, D.; Gvasalia, G.; Alavidze, Z.; Gogokhia, L.; Kuhl, S.; Abedon, S.T. Phage therapy in clinical practice: Treatment of human infections. *Curr. Pharm. Biotechnol.* 2010, 11, 69–86.
  110. NIH. New NIH Awards Will Support Development of Therapeutic Alternatives to Traditional Antibiotics.
  111. Chaplin, S. Topical antibacterial and antiviral agents: Prescribing and resistance. *Prescriber* 2016, 27, 29–36.
  112. Abedon, S.T.; Kuhl, S.J.; Blasdel, B.G.; Kutter, E.M. Phage treatment of human infections. *Bacteriophage* 2011, 1, 66–85.

113. Gu, J.; Liu, X.; Li, Y.; Han, W.; Lei, L.; Yang, Y.; Zhao, H.; Gao, Y.; Song, J.; Lu, R.; et al. A method for generation phage cocktail with great therapeutic potential. *PLoS ONE* 2012, 7, e31698.
114. Tothova, L.; Celec, P.; Babickova, J.; Gajdosova, J.; Al-Alami, H.; Kamodyova, N.; Drahovska, H.; Liptakova, A.; Turna, J.; Hodosy, J. Phage therapy of cronobacter-induced urinary tract infection in mice. *Med. Sci. Monit.* 2011, 17, BR173–BR178.
115. Dufour, N.; Clermont, O.; La Combe, B.; Messika, J.; Dion, S.; Khanna, V.; Denamur, E.; Ricard, J.D.; Debarbieux, L. Bacteriophage LM33\_P1, a fast-acting weapon against the pandemic ST131-O25b:H4 *Escherichia coli* clonal complex. *J. Antimicrob. Chemother.* 2016, 71, 3072–3080.
116. Laanto, E.; Bamford, J.K.; Ravantti, J.J.; Sundberg, L.R. The use of phage FCL-2 as an alternative to chemotherapy against columnaris disease in aquaculture. *Front. Microbiol.* 2015, 6, 829.
117. Kishor, C.; Mishra, R.R.; Saraf, S.K.; Kumar, M.; Srivastav, A.K.; Nath, G. Phage therapy of staphylococcal chronic osteomyelitis in experimental animal model. *Indian J. Med. Res.* 2016, 143, 87–94.
118. Gu, J.; Li, X.; Yang, M.; Du, C.; Cui, Z.; Gong, P.; Xia, F.; Song, J.; Zhang, L.; Li, J.; et al. Therapeutic effect of pseudomonas aeruginosa phage YH30 on mink hemorrhagic pneumonia. *Vet. Microbiol.* 2016, 190, 5–11.
119. Keen, E.C. Phage therapy: Concept to cure. *Front. Microbiol.* 2012, 3, 238.
120. Cooper, C.J.; Khan Mirzaei, M.; Nilsson, A.S. Adapting drug approval pathways for bacteriophage-based therapeutics. *Front. Microbiol.* 2016, 7, 1209.
121. Brayfield, A. *Martindale: The Complete Drug Reference*; Pharmaceutical Press: London, UK, 2014.
122. Allen, L.V. *Remington: An Introduction to Pharmacy*; Pharmaceutical Press: London, UK, 2013.
123. Aulton, M.E. *Aulton's Pharmaceutics: The Design and Manufacture of Medicine*; Churchill Livingstone: Edinburgh, IN, USA, 2007.
124. British Pharmacopoeia Commission. *British Pharmacopoeia*; Stationery Office: London, UK, 2012.
125. Alyami, H.; Dahmash, E.; Bowen, J.; Mohammed, A.R. An investigation into the effects of excipient particle size, blending techniques and processing parameters on the homogeneity and content uniformity of a blend containing low-dose model drug. *PLoS ONE* 2017, 12, e0178772.
126. Brown, T.L.; Thomas, T.; Odgers, J.; Petrovski, S.; Spark, M.J.; Tucci, J. Bacteriophage formulated into a range of semisolid and solid dosage forms maintain lytic capacity against isolated cutaneous and opportunistic oral bacteria. *J. Pharm. Pharmacol.* 2017, 69, 244–253.
127. O'Flaherty, S.; Ross, R.; Meaney, W.; Fitzgerald, G.; Elbreki, M.; Coffey, A. Potential of the polyvalent anti-staphylococcus bacteriophage K for control of antibiotic-resistant staphylococci from hospitals. *Appl. Environ. Microbiol.* 2005, 71, 1836–1842.
128. Chen, L.-K.; Liu, Y.-L.; Hu, A.; Chang, K.-C.; Lin, N.-T.; Lai, M.-J.; Tseng, C.-C. Potential of bacteriophage FAB2 as an environmental biocontrol agent for the control of multidrug-resistant *Acinetobacter baumannii*. *BMC Microbiol.* 2013, 13, 154.
129. Merabishvili, M.; Monserez, R.; van Belleghem, J.; Rose, T.; Jennes, S.; De Vos, D.; Verbeken, G.; Vanechoutte, M.; Pirnay, J.-P. Stability of bacteriophages in burn wound care products. *PLoS ONE* 2017, 12, e0182121.
130. Sansom, C. Phage therapy for severe infections tested in the first multicentre trial. *Lancet Infect. Dis.* 2015, 15, 1384–1385.
131. Brown, T.L.; Petrovski, S.; Dyson, Z.A.; Seviour, R.; Tucci, J. The formulation of bacteriophage in a semi solid preparation for control of *Propionibacterium acnes* growth. *PLoS ONE* 2016, 11, e0151184.
132. Biemer, J.J. Antimicrobial susceptibility testing by the kirby-bauer disc diffusion method. *Ann. Clin. Lab. Sci.* 1973, 3, 135–140.
133. Brown, T.L.; Petrovski, S.; Hoyle, D.; Chan, H.T.; Lock, P.; Tucci, J. Characterization and formulation into solid dosage forms of a novel bacteriophage lytic against klebsiella oxytoca. *PLoS ONE* 2017, 12, e0183510.
134. Brown, T.L.; Tucci, J.; Dyson, Z.A.; Lock, P.; Adda, C.G.; Petrovski, S. Dynamic interactions between prophages induce lysis in propionibacterium acnes. *Res. Microbiol.* 2017, 168, 103–112.
135. Todd, C. On the electrical behaviour of the bacteriophage. *Brit. J. Exp. Path.* 1927, 8, 369–376.
136. Serwer, P.; Pichler, M.E. Electrophoresis of bacteriophage T7 and T7 capsids in agarose gels. *J. Virol.* 1978, 28, 917–928.
137. Serwer, P.; Hayes, S.J. Agarose gel electrophoresis of bacteriophages and related particles. I. Avoidance of binding to the gel and recognizing of particles with packaged DNA. *Electrophoresis* 1982, 3, 76–80.
138. Cademartiri, R.; Anany, H.; Gross, I.; Bhayani, R.; Griffiths, M.; Brook, M.A. Immobilization of bacteriophages on modified silica particles. *Biomaterials* 2010, 31, 1904–1910.
139. Anany, H.; Chen, W.; Pelton, R.; Griffiths, M.W. Biocontrol of listeria monocytogenes and *Escherichia coli* O157:H7 in meat by using phages immobilized on modified cellulose membranes. *Appl. Environ. Microbiol.* 2011, 77, 6379–6387.

140. Piret, J.; Desormeaux, A.; Bergeron, M.G. Sodium lauryl sulfate, a microbicide effective against enveloped and nonenveloped viruses. *Curr. Drug Targets* 2002, 3, 17–30.
141. Sansom, L. *Australian Pharmaceutical Formulary and Handbook*, 23rd ed.; Pharmaceutical Society of Australia: Canberra, Australia, 2015; pp. 36–48.
142. Abedon, S.T.; Yin, J. Bacteriophage plaques: Theory and analysis. *Methods Mol. Biol.* 2009, 501, 161–174.
143. Thai, C.K.; Dai, H.; Sastry, M.S.; Sarikaya, M.; Schwartz, D.T.; Baneyx, F. Identification and characterization of cu(2)o- and zno-binding polypeptides by escherichia coli cell surface display: Toward an understanding of metal oxide binding. *Biotechnol. Bioeng.* 2004, 87, 129–137.
144. Kutateladze, M.; Adamia, R. Phage therapy experience at the eliava institute. *Med. Mal. Infect.* 2008, 38, 426–430.
145. Ryan, E.M.; Gorman, S.P.; Donnelly, R.F.; Gilmore, B.F. Recent advances in bacteriophage therapy: How delivery routes, formulation, concentration and timing influence the success of phage therapy. *J. Pharm. Pharmacol.* 2011, 63, 1253–1264.
146. Fish, R.; Kutter, E.; Wheat, G.; Blasdel, B.; Kutateladze, M.; Kuhl, S. Bacteriophage treatment of intransigent diabetic toe ulcers: A case series. *J. Wound Care* 2016, 25, S27–S33.
147. Lyu, X.; Zhao, C.; Yan, Z.M.; Hua, H. Efficacy of nystatin for the treatment of oral candidiasis: A systematic review and meta-analysis. *Drug Des. Devel. Ther.* 2016, 10, 1161–1171.
148. Miedzybrodzki, R.; Klak, M.; Jonczyk-Matysiak, E.; Bubak, B.; Wojcik, A.; Kaszowska, M.; Weber-Dabrowska, B.; Lobočka, M.; Gorski, A. Means to facilitate the overcoming of gastric juice barrier by a therapeutic bacteriophage a5/80. *Front. Microbiol.* 2017, 8, 467.
149. Netzer, P.; Gaia, C.; Sandoz, M.; Huluk, T.; Gut, A.; Halter, F.; Husler, J.; Inauen, W. Effect of repeated injection and continuous infusion of omeprazole and ranitidine on intragastric pH over 72 h. *Am. J. Gastroenterol.* 1999, 94, 351–357.
150. Tennant, S.M.; Hartland, E.L.; Phumoonna, T.; Lyras, D.; Rood, J.I.; Robins-Browne, R.M.; van Driel, I.R. Influence of gastric acid on susceptibility to infection with ingested bacterial pathogens. *Infect. Immun.* 2008, 76, 639–645.
151. Laheij, R.J.; Sturkenboom, M.C.; Hassing, R.J.; Dieleman, J.; Stricker, B.H.; Jansen, J.B. Risk of community-acquired pneumonia and use of gastric acid-suppressive drugs. *JAMA* 2004, 292, 1955–1960.