

Drug Development and Hepatotoxicity: A Comprehensive Exploration of Medical Advancement

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ABSTRACT

The research and manufacturing of novel compounds for medical use is necessary for the production of new drugs. Many medications with differing effects on health have been introduced throughout history. While some have proven advantageous, others have had unfavourable or even fatal consequences. This means that the process of introducing novel medications to the market is drawn out and entails extensive testing. First, medical problems and their symptoms are identified. Next, the therapeutic effects in cells and animal models are evaluated, which can have both favourable and negative outcomes. Peptide alterations' effects on tissue and cell structures, which in turn affect patient health, must be studied in order to gain a thorough understanding of medical disorders. Because the liver is located between the bloodstream and the gut, it plays a particularly important role in drug metabolism. Hepatotoxic chemicals are processed by it, and the resultant effects include pale stools, dark urine, stomach pain, frequent vomiting of blood, and exhaustion. Hepatotoxicity research frequently employs in vitro liver preparations and animal models to investigate the potential positive and negative effects of medications. Historical accounts detail the emergence of treatments for hepatitis, ranging from homeopathic cures to contemporary medications like nucleoside analogues and interferon. The development of safe and effective drugs requires a thorough understanding of liver function and drug interactions, underscoring the importance of thorough research in drug discovery and hepatotoxicity management.

Keyword: *Drug Development, Medical Treatment, Therapeutic Effects, Peptide Modifications, Liver Metabolism, Hepatotoxicity Animal Models, In Vitro Studies, Hepatitis Outbreaks, Interferon, Nucleoside Analogues, Drug Safety, Liver Function, Adverse Drug Reactions*

1. INTRODUCTION

The development and production of innovative substances that help in the medical management of health conditions is essential for the creation of new medications. There have been plenty of drugs released in history, a few of them possess beneficial effects on health, whereas some may have adverse, life-threatening, or just slight consequences on the functioning of the body. Consequently, the entry of these novel medicines into the marketplace demands a long time [1,2]. For an experimental medicine to qualify as appropriate for consumption, it has to pass via an extensive number of trials. Starting with the identification of the medical condition plus its manifestations that reduce the standard of life, novel remedies have been found. The toughest component when developing an innovative drug involves assessing its therapeutic effects in cells and experimental animals, that might result in positive as well as negative consequences. Therefore, these detrimental consequences can be thoroughly examined employing many different kinds of model animals. The most important step of figuring out the root cause of a medical condition is to understand how peptides undergo modifications, how this affects the structure of tissues and cells, and the way these

modifications influence well-being of patients [3,4]. The discovery of drugs isn't limited to recording beneficial outcomes; it is equally necessary to keep track of adverse reactions so as to avoid these research studies from having an impact on animals. Species such as rodents, guinea and additional species are utilized to conduct these experiments. It usually takes an extended period and a lot of effort to create drugs that are novel. Many medicinal products were eliminated throughout the last half-century, whereas hundreds of novel pharmaceuticals are currently on the market [1,2]. In the current time period, contaminants, chemicals, and industrial debris containing toxic substances are mixed together in the environment in which we live, especially the atmosphere and freshwater which is primarily needed by the bodies of humans to work efficiently. The organ known as the liver residing in animals. The liver is an organ that performs an integral part in the breakdown of medication thanks to its physical position that exists between the gut and the bloodstream [4,5]. The human liver is a versatile organ that manages both secretion and detoxifying tasks. It has a weight of approximately 1.5 kilograms in an adult male, which makes it one of the biggest glands in the human physique [6]. Once a drug develops a therapeutic response, it requires biological transformation in the cells of the liver. From intestinal absorption and portal venous circulation, an immense quantity of vitamins and minerals and foreign chemicals travel into the hepatocytes. Hepatotoxic substances such as kava, ephedra, and galactosamine are additionally metabolized by the liver's cells to hazardous byproducts [3].

The medical condition termed hepatotoxicity, which is additionally referred to as cirrhosis of the liver, is a condition that occurs when the functioning of the liver gets damaged by a wide range of conditions. These circumstances comprise foreign substances, encompassing pollutants from the environment, chemicals, drugs, dosages for therapeutic purposes, and chemical substances activity that has come into touch with the internal organs of a person. The liver purifies the human body against many different kinds of harmful substances that arise by means of chemical responses or when consuming any harmful substance. Like everyone will understand, the body of a person is capable of recognizing any strange occurrence, irrespective of how insignificant, severe, or dangerous it might seem. Symptoms of such disorders includes black urine, pale feces, cramping in the abdomen, blood vomit frequently, and fatigue. The smooth endoplasmic reticulum, formerly referred to as the metabolism clearance residence, exists in the liver's cells and serves as a catalyst for each intrinsic and exterior drug breakdown.

Omega-3 fatty acids, lipids, and steroids are converted to energy by endogenous procedures however prescription drugs gave for medicinal reasons or illegal drugs are broken down by external mechanisms. The mitochondria, a cellular organelle that generates ATP to support or accelerate the metabolic processes response, suffers damage through these chemical events. Abnormal levels of oxidizing agents are produced as a consequence of injury to mitochondria, that may cause damage to liver cells and specific metabolic enzymes like CYP2E1 which belongs to the cytochrome P-450 family. Bile duct cell lines might go through apoptosis primarily as a consequence to mitochondrial damage, that may cause additional damage the functioning of the liver. It may be possible for chemicals to negatively impact organs at the molecular or cellular level via complex interactions which take place at cellular levels & rely on either direct or indirect interactions with liver cells, that involve RNA, protein, and DNA [8]. Hepatic toxicity is researched utilizing a wide range of animal models- in vivo approach is not often employed, whereas the in vitro hepatic preparations has been used more often because it offers different viewpoints at every phase of investigate toxicity.

2. BACKGROUND

In 1957, interferon became known for the treatment of hepatotoxicity. Following that, many drugs and immunizations have also been developed for the same reason. Following then, PME anti-HBV activity was discovered in 1990. Hepatitis B and HIV treatment alongside 3TC anti-HBV and anti-HIV activity was first detected in 1991. Hepatitis B therapy using Interferon-Alfa 2B gained authorization in 1991. Entecavir's anti-HBV activity for hepatitis has been identified in 1998. Lamivudine (3TC), the very first nucleoside analogue of HBV, received FDA approval (1). Hepatitis was once an immense mystery, yet the enigma is about to be revealed. The first therapeutic text goes to the age of Sumerian's clay tablets, who mention jaundice in the 3rd millennium B.C. Devil Ahhazu (2) is the hepatitis-causing agent. Hippocrates disclosed the clinical manifestations of jaundice, which result in mortality in less than 12 days, from the years 375 and 460 B.C. Meal with water and honey was utilized as a remedy for this clinical symptom (3). First, there are several proofs of hepatitis which were propagated through Greeks and Romans but were erroneously identified for leptospirosis and malaria. Pope Zachary advised that the most effective remedy regarding this matter is isolation. During battles in the middle of the nineteenth century, especially the capture of Saint-Jean-d'Arcy in 1799 and the American Civil War (1861–1865), many cases of hepatitis circulated all over military groups. Hepatitis has been estimated had inflicted 16 million casualties during the course of World War II, including the US Army claiming 1,50,000 cases of the infection. Several epidemics were recorded during conflicts

in the 18th century, especially the Siege of Saint-Jean-Dacre in 1799 and the Siege of Paris in 1870. An additional problem during the American Civil War (1861–1865) was the 52 000 hepatitis cases. Hepatitis contributed a total of 16 million fatalities throughout World War 2. In comparison against the German military's 4 million "the census data," the US Army reported 150 000 cases (3). Those with immunizations against smallpox are also showing hepatitis symptoms. The writer and researcher of this study is Luhrman. A number of cases are additionally linked to the consumption of gold salt or bismuth iv and orally, accordingly. There were multiple cases from the US Navy in 1942, leading to 56000 patients getting the infection upon getting yellow fever vaccine contaminated with human plasma (4). Macallum knew the difference between the two kinds of hepatitis in 1947: epidemic hepatitis, that has a short time frame for incubation, and serum hepatitis, that requires an extended incubation period (100 days of fever) (5).

2.1 Hepatitis B virus became known via a big bang

While studying lipoprotein polymorphism at the prestigious National Institutes of Health (NIH) in 1963, Baruch Blumberg and his associates noticed a surprising relationship between Australian Aborigine oxygen-rich blood and poly-transfused hemophilic serum. Though the red staining of its reaction was different, he noticed that he had come across an unknown lipoprotein (6). Australian Antigen (Au), an innovative antigen, is responsible for the observed red staining. By irony, a lab technician acquired jaundice in 1967, and subsequent study reveals the presence of Au Antigen is linked to hepatitis (7). The serum antigen associated with serum Hepatic antigen (SH antigen) was identified for the first time in 1968 under a scientist by the name of Alfred Prince employing electrophoretic technique (8). The antigens that are responsible for Au and SH later turned out to be equivalent. The relationship that exists between post-trans fusional hepatitis and blood donors was proven in a study that took place in Japan in 1972. The previously determined antigen has been renamed as the HBs antigen, and its identification is consequently necessary. Fortunately, since the beginning of the twentieth century, nearly every one of notable organizations have gone through testing for blood centers. Blumberg developed the first generation HBs antigen plasma-based vaccination. Following all of this, he filed for a patent to safeguard his own developed version of the vaccination concept, and for the first time in history, a vaccine wasn't manufactured employing tissue culture. Later, in 1976, Blumberg bagged the Noble Price in Medicine for his groundbreaking research in manufacturing the HBV vaccination and characterizing HBV (9).

2.2 Discovery of Hepatitis virus A (HAV)-

NIH scientist Stephen Finestone established the immune-electron microscopy technique in the very early months of 1977 while partnering with Kalpakian to determine an agent in the stool containing the HAV antigen while looking for the rotavirus. The HAV antigen as well as its antibodies, IgG & IgM, produce themselves in the serum, as determined by serological investigations. A vaccine emerged quickly following tissue culture experiment (10).

2.3 Discovery of hepatitis delta virus (HDV)-

Employing immunostaining, a medical practitioner titled Mario Rizzuto identified a novel antigen in 1977 while attempting to identify HBV antigen in an Italian patient. This antigen is HDV. Although it is unlike HBE and HBC antigens, this antigen has been named as delta. The medical sign of the infectious disease comes due to the delta antigen, which is transported by HBV (11). A new flawed RNA virus which requires the HBV helper antigen for replication came to light in a study performed at the NIH because of scientists John Gerin and Robert Purcell who collaborated on transmission studies on chimpanzees. The HDV RNA virus was successfully identified; it is the tiniest vector to date (12). In certain regions of the Amazon Forests as well as central Africa, this disease-causing HDV viral spreads as an endemic infection (13). Interferon Alpha, a substance that inhibits the is prenylation stage needed to prevent HDV replication, is the antiviral therapy employed for the viral infection. The most effective method of therapy to date is the HBV vaccination.

2.4 Identification of Hepatitis C Virus (HCV)-

Post-trans fusional hepatitis showed up in a study carried out at the NIH by Finestone, Purcell, and Prince in a laboratory in New York. It has been triggered by a yet-unidentified agent, and researchers are optimistic that one day they will come across this new pathogen (14,15). Experts have been trying to inject this infectious virus using post-trans fusional hepatitis victims into Chimpanzee mammals. Infectious hepatitis is highlighted by extremely contagious Antihemophilic factors VII and IX. This post-trans fusional hepatitis virus features an endoplasmic

reticulum tubule form and an additional lipid bilayer (16). This enclosed virus HCV has a molecular circumference of 45-60 nm. For many years, experts have been trying to figure out its culture and microbiological description. Then, at the CDC, Danial Bradley and Michael Houghton's analytical teams cooperated to discover the deadly Hepatitis C Virus (HCV). This finding assists in the detection of novel neglected viruses that include hepatitis G virus and blood products-transmitted virus (17).

3. Epidemiology of hepatotoxicity

Recently published information proved that the frequency of drug-induced liver injury (DILI) fluctuated between 1 per 10,000 to 1 per 100,000 (18-29). Many current studies indicate an increased incidence of toxicity to the liver. A number of Western (30-33) and Asian (34) nations generated helpful data on hepatotoxicity diagnostics and medical treatment. According to epidemiological research, Iceland shows a yearly rate of 19.1 incidents for every 100,000 inhabitants (33), while France shows a yearly rate of 13.9 incidents for every 100,000 people, including 12% of them admitted and an overall mortality rate of 6% (31). Majority liver harm-causing drugs are antibiotics, including 46% of incidents of DILI documented by the USA DILI web (35). In accordance to an Italian case-control investigation that was backed by Swedish, English, and population-based research, the prevalence of drug-induced liver injury (DILI) were 4.1 peoples for every 100,000 peoples, and 50% of the individuals harmed consumed NSAIDS (31–33). With respect to the patient category (inpatients or outpatients) & the selection time frame, ranging from 2 to 10 years (36), outcomes from multiple research methods fluctuate considerably.

The overwhelming majority of studies claim that throughout India, 58% of DILI and liver damage instances involved anti-tubercular pharmaceuticals and 11% of cases featured anti-epileptic drugs. The fatality rate associated of anti-tubercular pharmaceuticals is double that among alternative existing pharmaceuticals (37), despite this percentage fluctuates with nations (38), considering every nation offers independent ways of recommending the medication, and certain people will make use of it whereas others will not. Based on one research project carried out by clinicians in the British Isles (U.K.) from 1994 and 1999, the overall rate of injury to the liver among people consuming isoniazid is 100 for every 100,000 instances, whereas for amoxicillin-clavulanic acid was 10 for every 100,000 instances (39). The most vulnerable age categories are older people and kids, compared to young people and adults, considering their resistance to infection, suitability, and adherence to pharmaceuticals are somewhat minimal (40,41). Pregnant women are considerably more vulnerable to liver damage than men simply because of anatomy-related parameters. Higher-income individuals tend to be more likely to suffer liver failure than mid-class or poor people because consumption of additional prescription drugs such as antibiotics, steroidal products, and supplements (42).

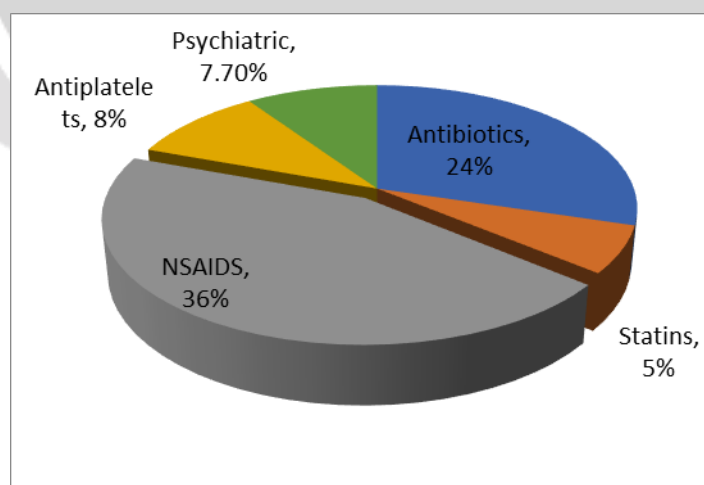


Fig -1 Percentage of instances involving drug-induced hepatotoxicity.

3.1 Cases of Hepatotoxicity based on the age-

The primary contributor to liver damage is the repeated use of pharmaceuticals like Amoxicillin-Clavulanate (46), halothane (47), isoniazid (48), and nitrofurantoin (49). Adolescent or adults may have hepatocellular liver toxicity,

however senior individuals are far more likely to suffer from cholestatic hepatotoxicity (50,51,52,53). Age isn't a big factor for certain medications (53). Hepatic toxicity with the antibiotic amoxicillin clavulanate is generally impacts adult and rarely impacts young people (55,56). Research (57, 56) indicate that taking antiepileptic medicines frequently among children promotes a rise in liver damage events.

3.2 Cases of Hepatotoxicity among Hospitalized Patients-

Only 17 out of 96 hospitalized patients (particularly older patients) who participated in Icelandic research display symptoms of liver damage, that can be brought upon through the consumption of multiple prescription drugs during the patient is at the medical center (57). Hepatic toxicity is extremely uncommon in as outpatients. Among approximately 1950 patients hospitalized to UK hospitals with an incidence rate of only 0.7%, there are just 13 individuals who display this liver damage manifestation (58). The prevalence rate of liver toxicity was 1.4% (59).

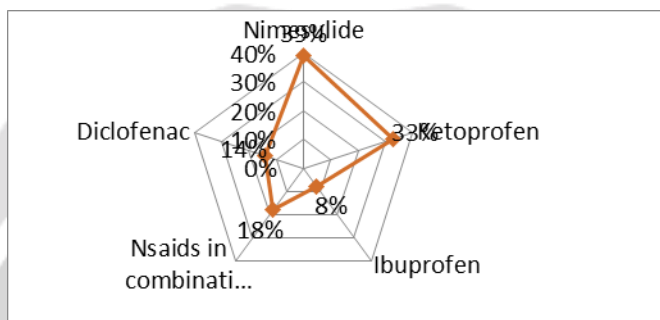


Fig-2 (RADAR) - This radar displays the 36% of instances in the cohort study analysis of the population that were caused by NSAIDS medicines.

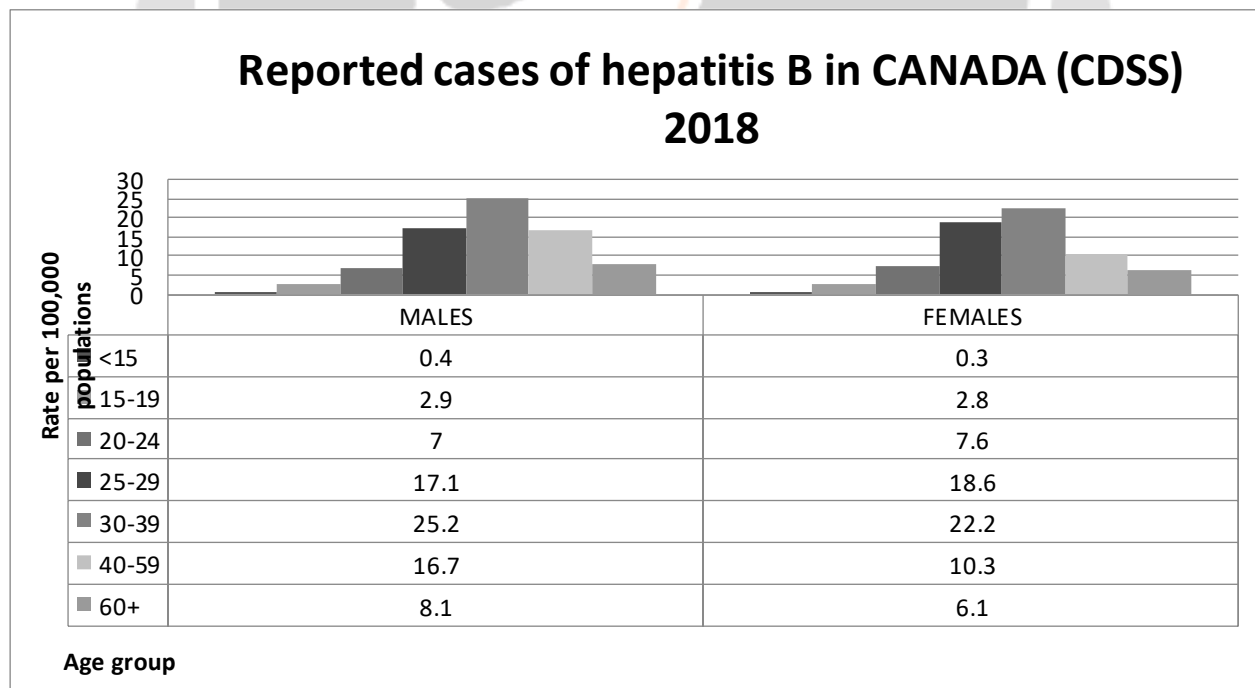


Fig-3 Rates of reported cases of chronic HBV infection by sex and age group in Canada, CNDSS, 2018 - Text description

4. Mechanism of action

Carbon tetra chloride (CCl₄)

CCl₄ is life-threatening dependent on the quantity and duration of contact. Consequences that include Ca⁺ shortage, peroxidation of lipids, release of cytokines, and programmed cellular death (Apoptosis) may take place at low concentrations. Cancer, fibrosis, and encephalopathy have been created when exposed to elevated quantities [1]. During the biological transformation of CCl₄ via cytochrome P450, trichloromethyl radical is generated [2]. After linking up with the unsaturated fatty acids found in the cell membrane, trichloromethyl radicals, considered harmful by its very nature, promote lipid peroxidation via the cross-linking of unsaturated fatty acids [3]

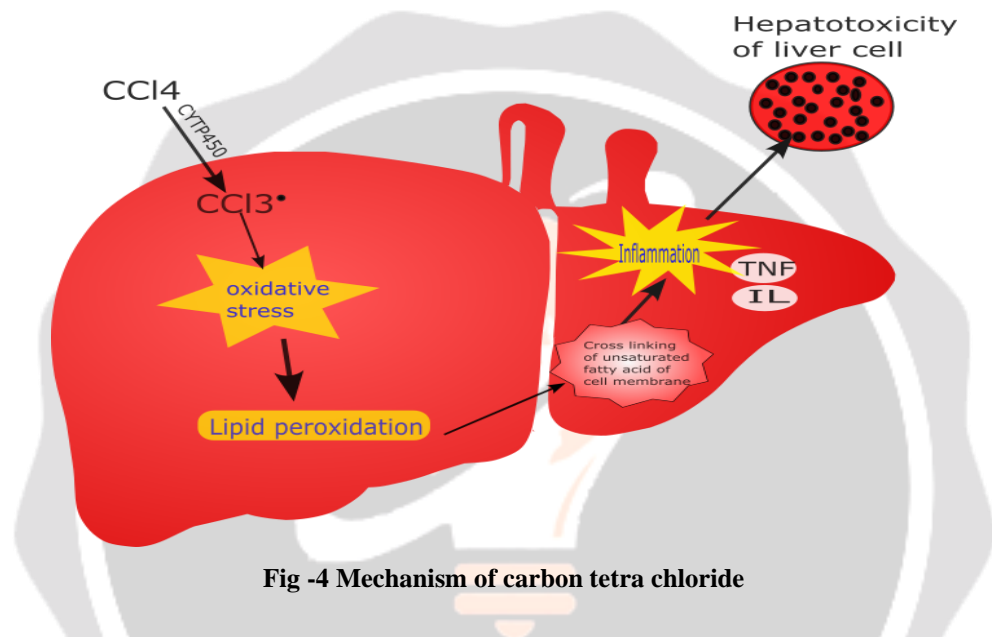


Fig -4 Mechanism of carbon tetra chloride

Isoniazid

For the medical management of tuberculosis, an isoniazid is employed. Although it displays more effectiveness & less catastrophic complications, the drug is sometimes considered to be a primary medicine. N-acetyltransferase (NAT2) is an enzyme likely helps in the breakdown of isoniazid. It converts the isoniazid directly into acetyl isoniazid, which is where acetyl hydrazine is generated via hydrolysis [4]. Acetyl hydrazine along with other intermediates efficiently adhere to big complexes like as proteins, lipids, DNA, and carbs [5], leading to cirrhosis and liver damage.

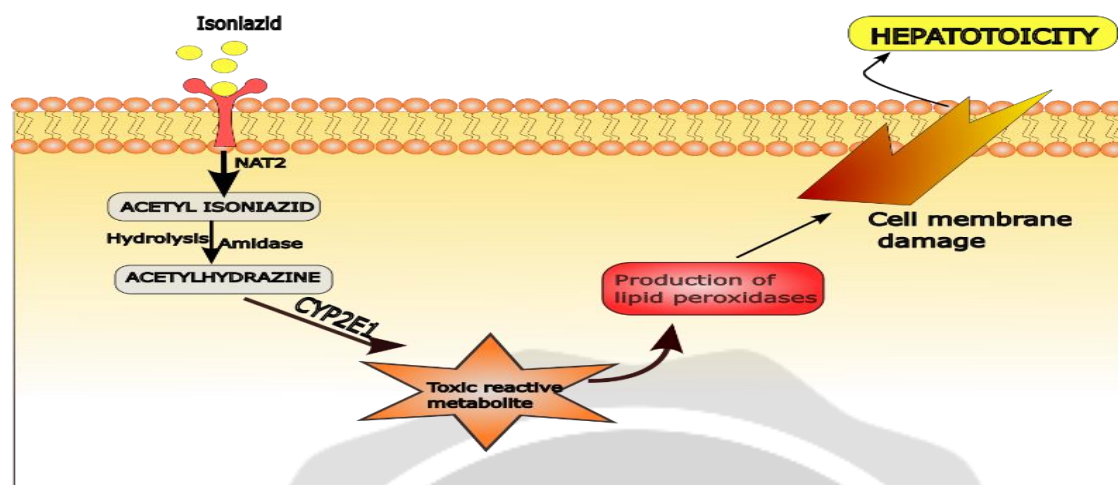


Fig-5 Mechanism of Isoniazid

Ethanol

Acetaldehyde gets created soon after the biological transformation of ethanol in with the help of (ADH) alcohol dehydrogenase [7]. Usually quite a bit quantity of ethanol is broken down through the pathways of ADH, MEOS, or the microbiota of the gut. However, when it comes a case of alcoholic’s damage to the liver, this alcohol dehydrogenase enzyme functions immediately. This refers to a condition during which ADH accelerates the biological transformation from ethanol towards acetaldehyde. If the disorder becomes more severe, mitochondrial failure will take place [8].

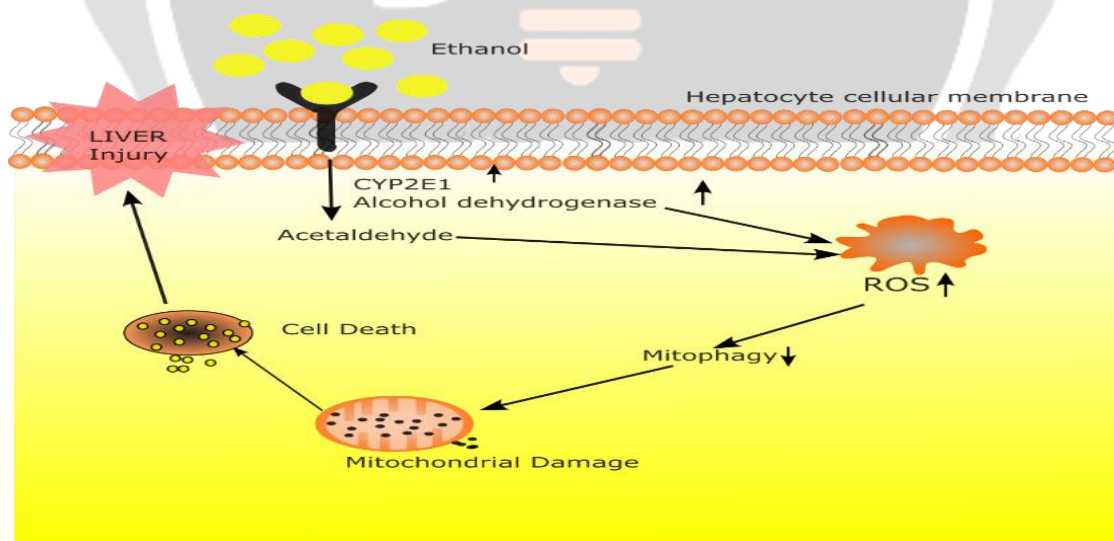


Fig-6 Mechanism of Ethanol

Methimazole

It belongs into the thioamides class and is employed for the management of hypothyroidism disorder [9]. the CYP2A6 enzyme, that processes down methimazole. It transforms methimazole to N-methylthiourea, which leads to cholestatic hepatitis [10, 11].

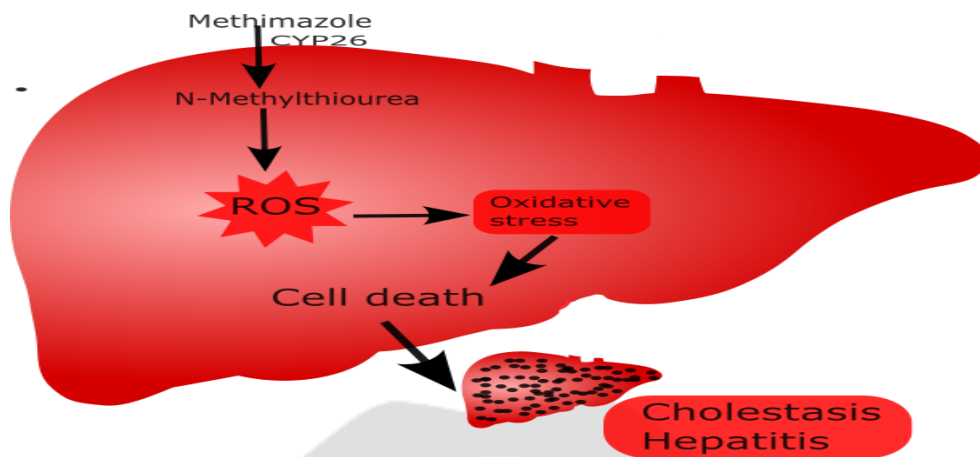


Fig-7 Mechanism of Methimazole

5. Hepatotoxicity Models

Carbon tetra chloride hepatotoxicity model

Rats from the Sprague Dawley/Albino strain were weighed (105–200g) [12] and separated into five groups. For group 1 rats, ordinary saline was used. When 75,90,105,120mg/kg of ccl4 were administered intramuscularly to group 2,3,4,5, lipid peroxidation and hepatotoxicity were witnessed. After 48, 72, 144 hours, one rat was removed from each group and sacrificed through their beheading [13].

Alcohol induced hepatotoxicity

Two groups of six rats each, Group A and Group B, were created from a total of twelve rats. Male albino rats weighing 150-200 g were weighed. Group B received treatment while Group A served as the control. For eight weeks, Group B (the treatment group) received 0.6ml/100gm/day of ethanol whereas Group A (the control group) received normal saline. Alcohol triggered hepatotoxicity in albino rats after 8 weeks (a rise in enzyme levels, mass rise in the liver, dysfunction of the mitochondria, fatty liver, etc.).[14]

Methimazole induced hepatotoxicity

Two groups (Group A and Group B) of 12 albino rats were formed based on weight. Group A rats received saline as the control group, and group B rats received 21 days of therapy with 60 mg/day of methimazole. After 21 days, the animals were sedated and slaughtered. Produced liver toxicity [15]

6. CONCLUSIONS

The development of new medications involves a detailed process of research and testing to ensure safety and efficacy. Historical data reveals that while some drugs have significantly advanced healthcare, others have posed risks, emphasizing the need for thorough evaluation. Understanding liver function and hepatotoxicity is crucial due to the liver's role in drug metabolism. Effective drug discovery relies on integrating findings from in vitro studies and animal models to identify potential risks. Ongoing research is essential to advance therapeutic options and enhance patient safety. Continued efforts in this field are vital for addressing challenges and improving drug safety.

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8. REFERENCES

1. 1.A textbook of Pharmaceutical regulatory science, By, Dr. R. Narayana Charyulu, Dr. Jobin Jose. NiraliPrakashan, Page No. .1.1-1.8.
2. A textbook of Pharmaceutical Regulatory science, By, Dr. Ashok Hajare, NiraliPrakashan, Page No,1.1.-1.36. www.google.com. A textbook of Pharmaceutical regulatory science, By, Dr. R. Narayana Charyulu, Dr. Jobin Jose. NiraliPrakashan, Page No. .1.1-1.8.
3. A textbook of Pharmaceutical regulatory science, By, Dr. R. Narayana Charyulu, Dr. Jobin Jose. NiraliPrakashan, Page No. .1.1-1.8.s
4. 3. A textbook of Pharmaceutical regulatory science, By, Dr. R. Narayana Charyulu, Dr. Jobin Jose. NiraliPrakashan, Page No. .1.1-1.8.
5. Steele VE, Boone CW, Lubet RA, Crowell JA, Holmes CA, Sigman CC, Kelloff GJ [1998], Preclinical drug development paradigms for chemopreventives. *HematolOncolClin North Am* 12;943-961, v-vi.
6. GDC Publication.
7. Timbrell JA [1983], Drug hepatotoxicity. *Br J ClinPharmacol* 15;3-14.
8. H. Gerhard Vogel *Drug Discovery and Evaluation Pharmacological Assays*, Second edition, Page no: 936-944.
9. N.S Parmar, Shiv Prakash “ Screening methods in pharmacology , Page no: 281-286.
10. Payen JL. De la jaunisse a l’hepatite C, 5000 ans d’histoire. Editions EDK: Paris, 2002.
11. Payen JL, Rongieres M. Histoire des hepatites. I. De la jaunisse au virus. *Rev Prat* 2002; 52: 2097–100
12. 3). Oon GCJ. Viral hepatitis - The silent killer. *Annals Academy of Medicine* 2012; 41: 279–80.
13. 4). Seeff LB, Beebe GW, Hoofnagle JH, et al. A serological follow-up of the 1942 epidemic of post-vaccination hepatitis in the United States Army. *N Eng J Med* 1987; 316: 965– 70.
14. 5). MacCallum F. Homologous serum hepatitis. *Lancet* 1947; 2: 691–2
15. 6). Alter HJ. The road not taken or how I learned to love the liver: a personal perspective on hepatitis history. *Hepatology* 2013; doi: 10.1002/hep.26787
16. 7). Blumberg BS, Alter HJ, Visnich A. A ‘new’ antigen in leukemia sera. *JAMA* 1965; 191:
17. 8. Prince A. An antigen detected in blood during the inoculation period of serum hepatitis. *Proc Natl Acad Sci USA* 1968; 60: 814–21.
18. 9). Blumberg BS. *Hepatitis B: The hunt for a killer virus*. New Jersey: Princeton University Press, 2003.
19. 10). Feinstone SM, Kapikian AZ, Purcell RH. Hepatitis A: detection by immune electron microscopy of virus like antigen associated with acute illness. *Science* 1973; 200: 365–73.
20. 11). Rizzetto M, Canese MG, Arico S, et al. Immunofluorescence detection of a new antigen/antibody system (Delta/ anti-Delta) associated with hepatitis B virus in liver and serum of HBsAg carriers. *Gut* 1977; 18: 997–1003.
21. 12). Purcell RH. The discovery of the hepatitis viruses. *Gastroenterology* 1993; 104: 955–63.
22. 13). Andrade ZA, Lesbordes JL, Ravisse P, et al. Fulminant hepatitis with microvesicular steatosis (a histologic comparison of cases occurring in Brazil – Labrea hepatitis – and in Central Africa – Bangui hepatitis. *Revista da Sociedade Brasileira de Medicina Tropical* 1992; 25: 155–60.
23. 14). Prince A, Brotman B, Grady G, et al. Long incubation post transfusion hepatitis without serological evidence of exposure to hepatitis B virus. *Lancet* 1974; 2: 2416.
24. 15). Feinstone S, Kapikian A, Purcell R, Alter H, Holland P. Transfusion associated hepatitis not due to viral hepatitis type A or B. *N Engl J Med* 1975; 292: 767–70.
25. 16). Bradley D, MacCaustland K, Cook E, et al. Post transfusion non A-non B hepatitis in chimpanzees; physicochemical evidence that the tubule-forming agent is a small, enveloped virus. *Gastroenterology* 1985; 88: 773–9.

26. 17). Choo Q, Kuo G, Weiner A, et al. Isolation of a cDNA clone derived from a blood-brone non A-non B viral hepatitis genome. *Sicence* 1989; 244: 359–62.
27. 18) Bjornsson ES. Drug-induced liver injury: an overview over the most critical compound. *ArchToxicol* 2015; 89: 327-334.
28. 19) Chalasani P, Hayashi P, Bonkovsky H, Navarro V, Lee W. ACG Clinical Guideline: the diagnosis and management of Idiosyncratic Drug-Induced Liver Injury. *Am J Gastroenterol* 2014; 109: 950-966.
29. 20) Navarro VJ, Senior JR. Drug related hepatotoxicity. *N Engl J Med* 2006; 354: 731-739.
30. 21) Reuben A, Koch DG, Lee WM. Drug induced acute liver failure: results of a US multicenter, prospective study. *Hepatology* 2010; 52: 2065-2076.
31. 22). Danan G. The Roussel Uclaf Causality Assessment Method (RUCAM) score is based upon a modification of the original scoring system of the Council for International Organizations of Medical Sciences (CIOMS) and supported by Roussel Uclaf Pharmaceuticals. *Consensus Meetings on: causality assessment of drug-induced liver injury. J Hepatol* 1988; 7: 132-136.
32. 23).Danan G, Benichou C. Causality assessment of adverse reaction to drugs--A novel method based on the conclusions of international consensus meeting: application to drug-induced liver injuries. *J Clin Epidemiol* 1993; 46: 1323-1330.
33. 24) Benichou, Danan G, Flahault A. Causality assessment of adverse reactions to drugs. An original model for validation of drug causality assessment methods: case reports with positive rechallenge. *J Clin Epidemiol* 1993; 46: 1331-1336.
34. 25) Maria VA, Victorino RM. Development and validation of a clinical scale for the diagnosis of drug induced hepatitis. *Hepatology* 1997; 26: 664-669.
35. 26) Aithal GP, Watkins PB, Andrade RJ, Larrey D, Molokhia M, Takikawa H, Hunt CM, Wilke RA, Avigan M, Kaplowitz N, Bjornsson E, Daly AK. Case definition and phenotype standardization in drug-induced liver injury. *Clin Pharmacol Ther* 2011; 89: 806-815.
36. 27) A. Licata. Adverse drugs reactions and organ damage: The liver. *Eur J Internal Med* 2016; 28: 9-16.
37. 28) Lucena MI, Andrade RJ, Fernández MC, Pachkoria K, Pelaez G, Durán JA, Villar M, Rodrigo L, Romero-Gomez M, Planas R, Barriocanal A, Costa J, Guarner C, Blanco S, Navarro JM, Pons F, Castiella A, Avila S; Spanish Group for the Study of Drug-Induced Liver Disease (Grupo de Estudio para las Hepatopatías Asociadas a Medicamentos (GEHAM)). Determinants of the clinical expression of amoxicillin-clavulanate hepatotoxicity: a prospective series from Spain. *Hepatology* 2006; 44: 850-856.
38. 29) Ahmad J, Odin J. Epidemiology and genetic Risk factors of drugs Hepatotoxicity. *Clin Liver Dis* 2017; 21: 55-72.
39. 30) Hoofnagle JH Drug induced liver injury Network (DILIN). *Hepatology* 2004; 40: 773.
40. 31) Sgro C, Clinard F, Ouazir K, Chanay H, Allard C, Guilleminet C, Lenoir C, Lemoine A, Hillon P. Incidence of drugs induced hepatic injuries: a French population-based study. *Hepatology* 2002; 36: 451-455.
41. 32). Andrade RJ, Lucena MI, Fernández MC, Pelaez G, Pachkoria K, García-Ruiz E, García-Muñoz B, González-Grande R, Pizarro A, Durán JA, Jiménez M, Rodrigo L, Romero-Gomez M, Navarro JM, Planas R, Costa J, Borrás A, Soler A, Salmerón J, Martín-Vivaldi R; Spanish Group for the Study of Drug-Induced Liver Disease. Drug-induced liver injury: an analysis of 461 incidences Submitted to Spanish registry over a 10 years period. *Gastroenterology* 2005; 129: 512-521.
42. 33). Bjornsson ES, Bergmann OM, Björnsson HK, Kvaran RB, Olafsson S. Incidence, presentation and outcomes in patients with drug-induced liver injury in the general population of Iceland. *Gastroenterology* 2013; 144: 1419-1425.
43. 34). Takikawa H, Murata Y, Horiike N, Fukui H, Onji M. Drug-induced liver injury in Japan: an analysis of 1676 cases between 1997 and 2006. *Hepato Res* 2009; 39: 427-431. A. Licata, M.G. Minissale, V. Calvaruso, A. Craxì 120.
44. 35). Chalasani N, Fontana RJ, Bonkovsky HI. Causes, clinical features, and outcomes from a prospective study of drug induced liver injury in the United States. *Gastroenterology* 2008; 135: 1924-1934.
45. 36). Donati M, Conforti A, Lenti M, Capuano A, Bortolami O. Risk of acute and serious liver injury associated to nimesulide and other NSAIDs: data from drug-induced liver injury case-control study in Italy. *Br J Clin Pharmacol* 2016; 82.1: 238-248.
46. 37). Devarbhavi H, R Dierkhising, Kremers WK. Single center experience with drug induced liver injury from India: causes, outcome, prognosis, and predictors of mortality. *Am J Gastroenterol* 2010; 105: 2396-2404.
47. 38). Bjornsson E, Olsson R. Outcome and prognostic markers in severe drug induced liver disease. *Hepatology* 2005; 42: 481-489.

48. 39). Pérez Gutthann S, García Rodríguez LA. The increased at risk of hospitalizations for acute liver injury in a population with exposure to multiple drugs. *Epidemiology* 1993; 4: 496-501.
49. 40). Nobili A, Licata G, Salerno F, Pasina L, Tettamanti M, Franchi C, De Vittorio L, Marengoni A, Corrao S, Iorio A, Marcucci M, Mannucci PM; SIMI Investigators. Polypharmacy, length of hospital stay, and in-hospital mortality among elderly patients in internal medicine wards. The REPOSI study. *Eur J Clin Pharmacology* 2011; 5: 507-519.
50. 41). Mannucci PM, A. Nobili Reposi investigator. *Intern Emerg Med* 2014; 9: 723-734.
51. 42). Morisco F, Bruno R, Bugianesi E, Burra P, Calvaruso V. AISF position paper on liver disease and pregnancy. *Dig Liver Dis* 2015; 48: 120-137.
52. 43). Hoofnagle JH. Serodiagnosis of acute viral hepatitis. *Hepatology* 1983; 2:267-8
53. 44). Kumar A, Mehrotra R. Non A, Non B hepatitis: the major etiologic agent of acute sporadic viral hepatitis in infancy and childhood in North India (personal communication). *Hepatology Rapid Lit Rev* 1989; 19(1):4-6.
54. 45). KaplowitjN, Aw TK, Simon FR, StolzaA. Drug induced hepatotoxicity. *Ann Intern Med* 1986; 104:826-39.
55. 46). Lucena MI, Andrade RJ, Fernández MC, et al; Spanish Group for the Study of Drug-Induced Liver Disease (Grupo de Estudio para las Hepatopatías Asociadas a Medicamentos (GEHAM)). Determinants of the clinical expression of amoxicillin-clavulanate hepatotoxicity: a prospective series from Spain. *Hepatology* 2006; 44(4):850–856
56. 47). Stock JG, Strunin L. Unexplained hepatitis following halothane. *Anesthesiology* 1985;63(4):424–439
57. 48). Fountain FF, Tolley E, Chrisman CR, Self TH. Isoniazid hepatotoxicity associated with treatment of latent tuberculosis infection: a 7- year evaluation from a public health tuberculosis clinic. *Chest* 2005;128(1):116–123
58. 49). Stricker BH, Blok AP, Claas FH, Van Parys GE, Desmet VJ. Hepatic injury associated with the use of nitrofurans: a clinicopathological study of 52 reported cases. *Hepatology* 1988;8(3):599–606
59. 50). Olsson R, Wiholm BE, Sand C, Zettergren L, Hultcrantz R, Myrhed M. Liver damage from flucloxacillin, cloxacillin and dicloxacillin. *J Hepatol* 1992;15(1-2):154–161
59. 50). Björnsson ES, Bergmann OM, Björnsson HK, Kvaran RB, Olafsson S. Incidence, presentation, and outcomes in patients with druginduced liver injury in the general population of Iceland. *Gastroenterology* 2013;144(7):1419–1425, e1–e3, quiz e19–e20
60. 51). De Valle MB, Av Klinteberg V, Alem N, Olsson R, Björnsson E. Druginduced liver injury in a Swedish University hospital out-patient hepatology clinic. *Aliment Pharmacol Ther* 2006;24(8):1187–1195
61. 52). Andrade RJ, Lucena MI, Fernández MC, et al; Spanish Group for the Study of Drug-Induced Liver Disease. Drug-induced liver injury: an analysis of 461 incidences submitted to the Spanish registry over a 10-year period. *Gastroenterology* 2005;129(2):512–521
62. 53). Lucena MI, Andrade RJ, Kaplowitz N, et al; Spanish Group for the Study of Drug-Induced Liver Disease. Phenotypic characterization of idiosyncratic drug-induced liver injury: the influence of age and sex. *Hepatology* 2009;49(6):2001–2009
63. 54). Squires RH Jr, Shneider BL, Bucuvalas J, et al. Acute liver failure in children: the first 348 patients in the pediatric acute liver failure study group. *J Pediatr* 2006;148(5):652–658
64. 55). Molleston JP, Fontana RJ, Lopez MJ, Kleiner DE, Gu J, Chalasani N; Drug-Induced Liver Injury Network. Characteristics of idiosyncratic drug-induced liver injury in children: results from the DILIN prospective study. *J Pediatr Gastroenterol Nutr* 2011;53(2): 182–189
65. 56).Devarbhavi H, Karanth D, Prasanna KS, Adarsh CK, Patil M. DrugInduced liver injury with hypersensitivity features has a better outcome: a single-center experience of 39 children and adolescents. *Hepatology* 2011;54(4):1344–1350.
66. 57). Björnsson ES, Bergmann OM, Björnsson HK, Kvaran RB, Olafsson S. Incidence, presentation, and outcomes in patients with druginduced liver injury in the general population of Iceland. *Gastroenterology* 2013;144(7):1419–1425, e1–e3, quiz e19–e20
67. 58). Duh MS, Walker AM, Kronlund KH Jr. Descriptive epidemiology of acute liver enzyme abnormalities in the general population of central Massachusetts. *Pharmacoepidemiol Drug Saf* 1999;8(4): 275–283
68. 59). Meier Y, Cavallaro M, Roos M, et al. Incidence of drug-induced liver injury in medical inpatients. *Eur J Clin Pharmacol* 2005;61(2): 135–

69. A textbook of Pharmaceutical regulatory science, By, Dr. R. Narayana Charyulu, Dr. Jobin Jose. NiraliPrakashan, Page No. .1.1-1.8.
70. A textbook of Pharmaceutical Regulatory science, By, Dr. Ashok Hajare, NiraliPrakashan, Page No,1.1.-1.36.
71. www.google.com.
72. GootzTD[1990], Discovery and development of new antimicrobial agents, ClinMicrobiol Rev 3;13-31.
73. Steele VE, Boone CW, Lubet RA, Crowell JA, Holmes CA, Sigman CC, Kelloff GJ [1998], Preclinical drug development paradigms for chemopreventives. Hemato/OncolClin North Am 12;943-961, v-vi.
74. GDC Publication.
75. Timbrell JA [1983], Drug hepatotoxicity. Br J ClinPharmacol 15;3-14.
76. A textbook of Pharmaceutical regulatory science, By, Dr. R. Narayana Charyulu, Dr. Jobin Jose. NiraliPrakashan, Page No. .1.1-1.8.
77. A textbook of Pharmaceutical Regulatory science, By, Dr. Ashok Hajare, NiraliPrakashan, Page No,1.1.-1.36.
78. www.google.com.
79. GootzTD[1990], Discovery and development of new antimicrobial agents, ClinMicrobiol Rev 3;13-31.
80. Steele VE, Boone CW, Lubet RA, Crowell JA, Holmes CA, Sigman CC, Kelloff GJ [1998], Preclinical drug development paradigms for chemopreventives. Hemato/OncolClin North Am 12;943-961, v-vi.
81. GDC Publication.
82. Timbrell JA [1983], Drug hepatotoxicity. Br J ClinPharmacol 15;3-14.
83. Delgado-Montemayor, Cecilia, et al. "Models of hepatoprotective activity assessment." *Medicina universitaria* 17.69 (2015): 222-228.
84. Johnston, David E., and Christine Kroening. "Mechanism of early carbon tetrachloride toxicity in cultured rat hepatocytes." *Pharmacology & toxicology* 83.6 (1998): 231-239.
85. Berger, Marc L., et al. "CCl₄- induced toxicity in isolated hepatocytes: The importance of direct solvent injury." *Hepatology* 6.1 (1986): 36-45.
86. Maddrey, Willis C. "Drug-induced hepatotoxicity: 2005." *Journal of clinical gastroenterology* 39.4 (2005): S83-S89.
87. Lei, Saifei, Ruizhi Gu, and Xiaochao Ma. "Clinical perspectives of isoniazid-induced liver injury." *Liver Research* 5.2 (2021): 45-52.
88. Walubo, A., P. Smith, and P. I. Folb. "The role of oxygen free radicals in isoniazid-induced hepatotoxicity." *Methods and findings in experimental and clinical pharmacology* 20.8 (1998): 649-656.
- Zakhari, S., & Li, T. K. (2007). Determinants of alcohol use and abuse: impact of quantity and frequency patterns on liver disease. *Hepatology*, 46(6), 2032-2039.
89. Méndez-Sánchez, Nahum, et al. "The mechanism of dysbiosis in alcoholic liver disease leading to liver cancer." *Hepatoma research* 6 (2020).
90. Li, Jianhua, et al. "Role of CYP2A6 in methimazole bioactivation and hepatotoxicity." *Chemical research in toxicology* 34.12 (2021): 2534-2539.
91. Singh, Gauri, and Ricardo Correa. "Methimazole." (2019).
92. Kobayashi, Masanori, et al. "Th2 cytokine-mediated methimazole-induced acute liver injury in mice." *Journal of applied toxicology* 32.10 (2012): 823-833.
93. Shah, Gopi H., BHARAT G. PATEL, and GAURANG B. SHAH. "Development of carbon tetrachloride-induced chronic hepatotoxicity model in rats and its application in evaluation of hepatoprotective activity of silymarin." *Asian journal of pharmaceutical and clinical research* 10.8 (2017).
94. Alhassan, A. J., et al. "Ideal hepatotoxicity model in rats using Carbon Tetrachloride (CCl₄)." *Bayero journal of pure and applied sciences* 2.2 (2009): 185-18.